Immunohistochemical Analysis of Metalloproteases in Dentigerous Cysts, Radicular Cysts and Keratocystic Odontogenic Tumors: Systematic Review

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ABSTRACT
Evidence suggests that metalloprotease expression may affect the biological behavior of odontogenic lesions. This study was conducted to review the literature about the role of metalloproteases in the development of odontogenic lesions. A search was carried out using one database, MEDLINE, via PubMed. Only articles written in English were included. Abstracts of all articles retrieved in the electronic search were evaluated for their relevance. Three articles met inclusion criteria. They analyzed the role of MMP-2, MMP-8 and MMP-13 in radicular cysts, dentigerous cysts and keratocystic odontogenic tumors, and of MMP-1, MMP-7 and MMP-27 in keratocystic odontogenic tumors. The immunostaining technique used for all studies was similar, differing only in type of staining used. Different immunoreactivity results were found in the studies. The pattern of metalloprotease expression in odontogenic lesions was different from the pattern found in other lesions. In the studies analyzed, there was a significant positive immunoreactivity for metalloproteases in odontogenic lesions, particularly in keratocystic odontogenic tumors, a finding that may explain KCOT aggressiveness.

Keywords: matrix metalloproteases, extracellular matrix, odontogenic lesions.

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Análise Imunoistoquímica de Metaloproteases nos Cistos Dentígeros, Cistos Radiculares e Tumores Odontogênicos Ceratocísticos: Revisão Sistemática

RESUMO

Evidências sugerem que a expressão das metaloproteinases podem afetar o comportamento biológico das lesões odontogênicas. Esse estudo foi conduzido a fim de revisar a literatura sobre o papel das metaloproteinases no desenvolvimento das lesões odontogênicas. A pesquisa foi realizada utilizando a base de dados do MEDLINE, via PUBMED. Somente artigos escritos em língua inglesa foram aceitos. Os resumos de todos os artigos encontrados na busca foram avaliados de acordo com sua relevância. Três artigos preencheram os critérios de inclusão. Eles analisaram o papel da MMP-2, MMP-8 e MMP-13 nos cistos radiculares, cistos dentígeros e nos tumores odontogênicos ceratocísticos (TOC) e MMP-1, MMP-7 e MMP-27 no TOC. A técnica imunoistoquímica utilizada por todos os estudos foi similar, diferindo somente pelo tipo da coloração utilizada. Diferentes imunomarcações foram encontradas nos estudos. O padrão da expressão das metaloproteinases nas lesões odontogênicas variou entre as lesões. Nos estudos analisados, houve uma imunomarcação positiva, significante estatisticamente, das metaloproteinases nas lesões odontogênicas em especial nos TOCs, o que pode explicar a agressividade dessas lesões.

Palavras-chave: metaloproteinases da matriz, matriz extracelular, lesões odontogênicas.

INTRODUCTION

The molecules of the extracellular matrix play an important role during cell development and morphogenesis. Matrix metalloproteases (MMPs) are zinc-dependent proteinases that participate in extracellular matrix degradation (1-3) and favor the invasion and proliferation of tumor cells (4). Under normal physiological condition, MMPs are weakly expressed in tissues, whereas in pathological events, their overexpression is the cause of the imbalance between MMPs and tissue inhibitors of MMPs (TIMPs) (5-6-7).

To this date, 28 genes of the metalloprotease family have been identified in human beings. They are classified into 5 groups according to substrate specificity and internal homology: collagenases, gelatinases, stromelysins, membrane-type MMPs, and others, including matrilysins (8).

Some studies about odontogenic cysts and tumors have been conducted to evaluate metalloprotease expression in these lesions. MMP-1, MMP-2, MMP-3, MMP-8, MMP-9 and MMP-13 have been identified in odontogenic cysts, which suggests that MMPs play an important role in the growth of these tumors (9-10).

MMP-2 and MMP-9 are gelatinases of the MMP family that degrade some types of collagen, such as type IV, V and X collagen, and denature type I, II and III fibrillar collagen. They have been found on the walls of odontogenic cysts and in cystic fluid. In addition, they seem to be involved in the pathologic process of cyst expansion. Both MMP-2 and MMP-9 are secreted in their inactive form and have to be converted into their active form to perform their function. Studies found that the MMP-9 activity rate is directly associated with the type of odontogenic cyst (9-11).
MMP- and MMP-13, also known as collagenases, may directly initiate a critical breakdown of collagen, both in benign and malignant bone-destructive lesions (12).

Other studies also correlate matrilysins (MMP-7 and MMP-26) with lesions of odontogenic origin, particularly keratocystic odontogenic tumors (KCOT) (13). Matrilysins seem to degrade the substrate of the basement membrane and are correlated with cell proliferation, apoptosis, invasion and metastases.

Because of the importance of MMPs in the regulation, integrity and composition of the extracellular matrix and the possible correlation of MMPs with pathological processes, this study reviewed findings in the literature about this topic to improve our understanding about the biological processes involved in the development of odontogenic lesions.

METHODS

Review strategy

The studies were selected using the MEDLINE database (via PubMed). Only studies published between 1990 and 2009 were included. The following keywords were used: (metalloproteinase “OR” metalloproteinases “OR” metalloprotease “OR” metalloproteases “OR” MMP “OR” MMPs) AND odontogenic AND (cyst “OR” cysts “OR” keratocyst “OR” keratocysts). After the list of references was retrieved, only studies published in English were included in the review.

Criteria for sample selection

The title and abstract of all studies retrieved in the electronic search were evaluated according to their adequacy. The full texts of the studies selected were reviewed, and a decision was made about their eligibility for inclusion. To be eligible for data extraction, the studies had to meet the following criteria: 1) original research studies; 2) studies with immunohistochemical analyses; 3) inclusion of at least one type of metalloprotease in the study; and 4) inclusion of at least one of the three diseases under analysis (radicular cyst, dentigerous cyst, KCOT).

Data extraction

Of the studies selected, the following main data were extracted: 1) Country of origin; 2) Metalloproteases included; 3) Sample composition and size; 4) Methods used; and 5) Results found.
RESULTS

The MEDLINE search (via PubMed) yielded 15 potentially eligible studies. Their abstracts were selected according to their relevance. After screening, 11 studies remained for full text analysis. The four studies excluded in the first analysis (14-15-16-17) dealt with metalloprotease expression in lesions not evaluated in this study. After reading the full texts, 8 studies were classified as inadequate and excluded: 05 (11-18-19-20-21) had no immunohistochemical analyses; 02 (22-23) did not involve metalloproteases directly; and 01 (24) was a literature review. Therefore, three studies met inclusion criteria and were analyzed for data extraction (Graph 1).

Of the studies analyzed, two were conducted in Finland and one, in Brazil. Mean number of authors per study was 08. The metalloproteases, the lesion evaluated in each study and the sample size are shown in Table 1. MMP-2, MMP-8 and MMP-13 were
analyzed in radicular cysts, dentigerous cysts and KCOT. In addition, MMP-1, MMP-7 and MMP-27 were analyzed in KCOTs.

The methods used for the immunohistochemical analysis in each study are shown in Table 2, which describes antibody, dilution, antigen retrieval, incubation, staining and counterstaining. The method used in all studies was similar and only staining was different.

<table>
<thead>
<tr>
<th>Study</th>
<th>Metalloproteases</th>
<th>Diseases</th>
<th>Sample Size</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wahlgren et al., 2001</td>
<td>MMP-8, MMP-13</td>
<td>Periapical granulomas</td>
<td>10</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-2, MMP-8, MMP-13</td>
<td>Dentigerous cysts</td>
<td>10</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-2, MMP-8, MMP-13</td>
<td>Dentigerous cysts</td>
<td>14</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMMP-1, MMP-7, MMP-27</td>
<td>KCOT</td>
<td>11</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-1, MMP-13</td>
<td>KCOCT</td>
<td>11</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-8, MMP-13</td>
<td>Myelomas</td>
<td>07</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-2, MMP-8, MMP-13</td>
<td>Myelomas</td>
<td>07</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-1, MMP-7, MMP-27</td>
<td>Syndromic KCOT</td>
<td>16</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-1, MMP-7, MMP-27</td>
<td>Syndromic KCOT</td>
<td>20</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-1, MMP-7, MMP-27</td>
<td>Syndromic KCOT</td>
<td>21</td>
</tr>
</tbody>
</table>

KCOT: keratocystic odontogenic tumor.

<table>
<thead>
<tr>
<th>Study</th>
<th>Antibody</th>
<th>Dilution</th>
<th>Antigen retrieval</th>
<th>Incubation</th>
<th>Staining</th>
<th>Counterstaining:</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wahlgren et al., 2001</td>
<td>MMP-8</td>
<td>1:800</td>
<td>Pepsin – 45min</td>
<td>Overnight</td>
<td>3-amino-9-ethylcarbazole</td>
<td>Mayer’s hematoxylin</td>
</tr>
<tr>
<td>Wahlgren et al., 2001</td>
<td>MMP-13</td>
<td>1:1000</td>
<td>Pepsin – 45min</td>
<td>Overnight</td>
<td>3-amino-9-ethylcarbazole</td>
<td>Mayer’s hematoxylin</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-2</td>
<td>1:1000</td>
<td>Pepsin – 60min</td>
<td>Overnight</td>
<td>Mayer’s hematoxylin</td>
<td>alcine</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-8</td>
<td>1:500</td>
<td>Pepsin – 60min</td>
<td>Overnight</td>
<td>Mayer’s hematoxylin</td>
<td>alcine</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-13</td>
<td>1:1000</td>
<td>Pepsin – 60min</td>
<td>Overnight</td>
<td>Mayer’s hematoxylin</td>
<td>alcine</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-7</td>
<td>1:250</td>
<td>Citrate - 30min</td>
<td>Overnight</td>
<td>Mayer’s hematoxylin</td>
<td>alcine</td>
</tr>
<tr>
<td>Wahlgren et al., 2003</td>
<td>MMP-26</td>
<td>1:250</td>
<td>Pepsin – 60 min</td>
<td>Overnight</td>
<td>Mayer’s hematoxylin</td>
<td>alcine</td>
</tr>
</tbody>
</table>
The methods to analyze the antibodies expressed in each study are shown in Table 3. Three different methods were used to evaluate immunoexpression, and accuracy increased with publishing date.

**TABLE 3 – Method to analyze antibody expression in each study.**

<table>
<thead>
<tr>
<th>Study</th>
<th>Immunoreactivity Analysis - Score Used</th>
</tr>
</thead>
</table>
| Wahlgren et al., 2001 | No immunoreactivity  
|                   | Positive immunoreactivity                                     |
| Wahlgren et al., 2003 | 0 – no immunoreactivity  
|                   | 1 – positive immunoreactivity  
|                   | 1 - Weak  
|                   | 2 – Moderate  
|                   | 3 – Strong immunoreactivity *                                   |
| Cavalcante et al., 2008 | - Negative  
|                   | + Positive  
|                   | ++ Strongly positive **                                        |

* Immunoreactivity evaluation according to Bachmeier et al. (modified technique)  
* Immunoreactivity evaluation according to Kumamoto et al.

Because of the difficulty in standardizing the analysis of immunoexpression, results were classified as: no immunoreactivity (-) or positive immunoreactivity (+). When immunoreactivity was present, its intensity was not analyzed. Results of immunoreactivity for each enzyme in the studies are summarized in Table 4. The pattern of metalloprotease expression in KCOTs is different from that found in other lesions.
TABLE 4 – Results of immunoreactivity for each enzyme in each study.

<table>
<thead>
<tr>
<th>Study</th>
<th>Dentigerous cysts</th>
<th>Radicular cysts</th>
<th>Keratocystic odontogenic tumors</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wahlgren et al., 2001 (plasma cells)</td>
<td>MMP-8</td>
<td>MMP-13</td>
<td>MMP-8</td>
</tr>
<tr>
<td></td>
<td>- 4</td>
<td>- 6</td>
<td>+ 6</td>
</tr>
<tr>
<td></td>
<td>40%</td>
<td>60%</td>
<td>75%</td>
</tr>
<tr>
<td>Wahlgren et al., 2003 (epithelial cells)</td>
<td>MMP-2</td>
<td>MMP-8</td>
<td>MMP-13</td>
</tr>
<tr>
<td></td>
<td>+ 2</td>
<td>- 8</td>
<td>+ 4</td>
</tr>
<tr>
<td></td>
<td>20%</td>
<td>80%</td>
<td>36%</td>
</tr>
<tr>
<td>Cavalcante et al., 2008 (epithelial cells)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>10%</td>
<td>90%</td>
<td>25%</td>
</tr>
</tbody>
</table>
DISCUSSION

The scarcity of studies that investigate the association between metalloproteases and lesions of odontogenic origin explains the small number of studies retrieved in this review. Although the search was applied in different ways, there were no changes in results. In general, very few studies have been conducted to evaluate the presence of these enzymes in other lesions, which may be explained by the fact that the discovery of these enzymes is recent. MMP-1 was first described by Jerome Gross and Charles Lapiere in 1962. They found enzymatic activity (collagen triple-helix degradation) during the metamorphosis of a tadpole tail (25). This enzyme was called interstitial collagenase. Currently, the descriptions of 28 different types of metalloproteases are found in the literature, and MMP-28 was only discovered in 2001 (26).

After the initial selection of studies (n=15), only 03 met the criteria for data extraction. Coincidently, two studies had been conducted by the same research team. In the final selection, the immunohistochemical evaluation of different cell types, plasma cells and epithelial cells, were also analyzed. Initially, the type of cell under analysis would be an exclusion criterion, and only the analysis of epithelial cells would be included. However, the inclusion of both types was possible because of the small number of studies about the topic of interest.

Several metalloproteases were evaluated in the studies selected. These enzymes are subdivided into six groups according to their substrate and to the similarity of their structural domain. The groups analyzed in this study were collagenases (MMP-1, MMP-8 and MMP-13), gelatinases (MMP-2) and matrilysins (MMP-7 and MMP-26). Collagenases are enzymes that can break the links that make up type I, II and III interstitial collagen, as well as other components of the extracellular matrix. Gelatinases can digest collagen after degradation (3). MMP-2 is directly associated with osteogenesis (27). Matrilysins are proteases that contribute to the degradation of the cell membrane and are involved in several processes, such as cell proliferation, apoptosis, invasion and metastases (13). All these enzymes are directly associated with physiological processes, but are overexpressed in pathological events.

Radicular cysts are classified as inflammatory cysts that originate from the epithelial cell rests of Malassez, secondary to pulp necrosis (28). Dentigerous cysts are developmental cysts, usually asymptomatic, that may potentially lead to cortical expansion and bone fenestration. They are associated with the crown of an impacted tooth. The exact pathogenesis of dentigerous cysts remains unknown, but several authors believe that they develop from tooth follicles (29). Some studies evaluated epithelial and mesenchymal characteristics of different cysts to elucidate questions that remain unclear in the literature. Some proteins and enzymes of the extracellular matrix seem to be associated with the development and biological behavior of these cysts. The variable patterns of metalloprotease expression in cyst epithelium indicate that they have different roles in regulating proliferation, maturation and cell migration. In the studies reviewed, immunoreactivity of dentigerous and radicular cysts for MMP-2, MMP-8 and MMP-13 was found in two different studies. MMP-2 (gelatinase) expression in dentigerous cysts
reached 80% immunoreactivity, whereas for MMP-8 and MMP-13, it was about 60% and 30%. The low immunoexpression of MMP-13, in both dentigerous and radicular cysts, may be associated with its role in bone-destructive lesions. MMP-13 is associated with the uncontrolled destruction of the extracellular matrix and of the bilaminar zone in aggressive malignant lesions (12).

KCOT is a benign tumor with high recurrence rates. According to several studies, recurrence ranges from 3% to 60% (30). Keratocysts were reclassified as keratocystic odontogenic tumors by the World Health Organization (WHO) due to their aggressive behavior and histology and new information about their genetics (31-32-33). The WHO defines them as benign uni- or multicystic tumors of odontogenic origin with a characteristic lining of parakeratinized stratified squamous epithelium and a potential for aggressive, infiltrative behavior.

New information about KCOT, particularly regarding bone resorption and growth, have led to changes in its treatment. Several recent studies focused on the collagenolytic activity of this tumor because this same activity is not found in other odontogenic lesions. Also, the activity of collagenase in tissues is strongly controlled by a complex regulating system that may have collagenolytic effects and, therefore, influence the expansion of the bone caused by the cyst (24) Some studies evaluated the presence, activity, activation and inhibition of extracellular matrix metalloproteases in KCOT to define the role of these enzymes in the molecular mechanisms associated with cystic growth. The study under analysis here also evaluated MMP-1, MMP-2, MMP-8, MMP-13 and MMP-27 in KCOTs. Strong immunoreactivity for these enzymes was found in all studies, except for MMP-8 in only one study. This may be explained by the fact that the analysis was conducted using plasma cells, and not epithelial cells. Differentiated plasma cells produce cytokines, such as interleukin-1, found in cystic fluid and responsible for MMP-9 and MMP-13 regulation (10). MMP-8 has a role in focal remodeling and cystic epithelial growth, but does not participate in the modulation of molecules in the bilaminar zone (12).

In the studies under analysis, there was a stronger expression of MMP-2 and MMP-13 in KCOTs than in dentigerous and radicular cysts. MMP-2 and MMP-13 are produced by the cystic epithelium and are found in the bilaminar zone. Some authors (10) suggest that these enzymes may induce epithelial migration due to the fragmentation of laminin-5 (transmembrane protein found exclusively in the bilaminar zone) and that this may explain the stimulation of migration and the growth potential of KCOTs. They also found that MMP-2 may activate latent MMP-13, which suggests that this enzyme may act as a cascade in the activation of epithelial migration, induced by fragments of degraded laminin. They concluded that the fast proliferation of the epithelium and the tendency to separate from the connective tissue capsule may be associated with laminin-5 modulation by MMP-2 and MMP-13 in KCOTs.

Immunoreactivity for MMP-1 was found in 90% of the cases, and it was the second strongest immunoreactivity in all studies under analysis. Type I collagen, responsible for the strength and rigidity of the connective tissue, is the main organic matrix of bone (34),
and MMP-1 is one of the proteases that may degrade the triple-helix domain of type I collagen (35-36). The presence of MMP-1 in KCOTs may be associated with the organic degradation of the bone matrix, which favors the dissemination of the disease through the trabecular spaces (13). According to some authors, it is still too early to suggest that a patient with strong MMP-1 immunoexpression should be warned about possible greater susceptibility to the development of the nevoid basal cell carcinoma syndrome (13). However, future studies may use immunoreactivity for some enzymes to clarify diagnoses and plan treatments.

Positivity was found for MMP-7 and MMP-27, the matrilysins, in 75% and 80% of the cases. These enzymes are associated with basement membrane degradation. In addition, they can activate other metalloproteases, such as MMP-2 and MMP-9 (gelatinases). These gelatinases degrade type IV collagen and basement membrane laminin (13). The role of matrilysins in odontogenic lesions has been evaluated only by the same research team, which precludes further scientific correlations.

CONCLUSIONS

• Further studies should be conducted to evaluate the correlations between metalloproteases and odontogenic lesions.

• In the studies analyzed, an important immunoreactivity for metalloproteases was found in odontogenic lesions, particularly in KCOTs.

• KCOTs have a more aggressive biological behavior, which may be currently explained by immunoreactivity for several proteins and enzymes, particularly metalloproteases.

REFERENCES


